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Research paper

Physical stability of redispersible dry emulsions containing amorphous sucrose

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Abstract

The objective of the present study was to estimate the stability of redispersible dry emulsions containing amorphous sucrose. Dry emulsions were prepared by spray drying liquid o/w-emulsions in a laboratory spray dryer. The effect of hydroxypropyl methylcellulose (HPMC) on the glass transition temperature $T_{\rm g}$ of spray dried sucrose-HPMC mixtures, relative to the $T_{\rm g}$ of amorphous sucrose, was investigated. For the sucrose-HPMC mixtures the values of $T_{\rm g}$ followed the ideal Gordon-Taylor equation up to 30% HPMC. For dry emulsions containing 40% HPMC, 30% lipid and 30% sucrose, the $T_{\rm g}$ was increased by 12°C relative to the $T_{\rm g}$ of amorphous sucrose. The stability of the dry emulsions was investigated by a conventional stability study and by an enthalpy relaxation study. The measured enthalpy recovery of amorphous sucrose below $T_{\rm g}$ was used to calculate molecular relaxation time parameters based on the Williams-Watts equation. The molecular mobility of amorphous sucrose at temperatures 50°C below $T_{\rm g}$ was low and negligible with respect to the shelf life stability. It was concluded that the dry emulsions are physically stable with respect to the lifetime of a pharmaceutical product when stored in dry condition and at temperatures up to 28°C. © 2002 Elsevier Science B.V. All rights reserved.

Keywords: Dry emulsion; Sucrose; Hydroxypropylmethylcellulose (HPMC); Glass transition temperature; Stability; Molecular mobility; Amorphous; Enthalpy relaxation

1. Introduction

Liquid emulsions have distinct advantages over other oral dosage forms by improving the bioavailability and by reducing the side effects, but the number of emulsion formulations currently in use are few compared with other oral dosage forms. The lack of usage is due to physical-chemical stability and compliance problems. Packages are often big and must be stored at low temperatures.

Dry emulsions are prepared by drying liquid o/w-emulsions containing a solid carrier in the aqueous phase. The solid carrier provides the dry emulsions with bulk and mass. Dry emulsions are lipid-based powder formulations from which an o/w-emulsion can be reconstituted in vivo [1–3] or in vitro [2,4–18].

From a pharmaceutical point of view, dry emulsions are attractive because they are physically and microbiologically stable solid formulations, which are easy to administer in the form of powders as capsules and tablets. Like o/w-emul-

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sions, dry emulsions represent a potential oral drug delivery system for lipophilic and low-soluble drug substances [1–3,6–9,12,15–18] and for drug substances needing protection against light [10] or oxidation [19].

Previous studies have shown that dry emulsions having a lipid content of up to 40% dry powder mass are physically stable [20]. These dry emulsions were prepared by spray drying liquid o/w-emulsions containing fractionated coconut oil dispersed in aqueous solutions of hydroxypropylmethylcellulose (HPMC). Unfortunately, the dry emulsions were cohesive powders having poor flow properties. The cohesiveness was reduced by addition of sucrose [21], meaning that the processing properties of the dry emulsions were improved.

Preparation of dry emulsions by spray drying transforms sucrose into an amorphous state, which exhibits a strong tendency to crystallise when stored at temperatures above the glass transition temperature $T_{\rm g}$. The relationship between $T_{\rm g}$ and the stability of amorphous solids is complex. The molecular mobility decreases as the temperature decreases, particularly below $T_{\rm g}$, thereby slowing molecular diffusion and reducing the rate of crystallisation. It is clear that storage below $T_{\rm g}$ under dry conditions is advantageous,

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though not a guarantee, for stability. Amorphous solids may be physically stabilised when combined with a polymer (an antiplasticizer) having a higher $T_{\rm g}$, making up an amorphous solid solution [22–24]. The combined system is expected to have a $T_{\rm g}$ between those of the individual components and a different degree of molecular mobility at a particular temperature and relative humidity [24].

From enthalpy relaxation studies, the molecular mobility can be estimated from the calculated molecular relaxation time parameters τ and β based on the Williams-Watts equation. This equation was originally developed to describe non-symmetrical dielectric relaxation behaviour and is frequently applied to quantify the relaxation process in amorphous solids [24–28]. Studies have shown that the molecular mobility becomes negligible from 50°C below $T_{\rm g}$, because the molecular mobility is too low to initiate crystallisation within the expected lifetime of amorphous pharmaceutical products [25,28–29].

The objective of the present study was to investigate the physical stability of redispersible dry emulsions containing amorphous sucrose and to find the temperature range at which the molecular mobility decreases to a level where it becomes negligible with respect to the expected lifetime of pharmaceutical products. The physical stability of the dry emulsions, the effect of HPMC on $T_{\rm g}$ and the molecular mobility of amorphous sucrose below $T_{\rm g}$ were investigated.

2. Materials and methods

2.1. Materials

HPMC (Pharmacoat® 606) Shin-Etsu (Japan) having a viscosity of 6.1 mPas (2% w/v solution) at 20°C, fractionated coconut oil (Miglyol® 812 N) Condea (Germany) and sucrose, Danisco Sugar (Denmark) were used.

2.2. Preparation of dry emulsions, amorphous sucrose and mixtures

Liquid o/w-emulsions with 20% dry powder mass were prepared so dry emulsions contained 40% HPMC, 30% fractionated coconut oil and 30% sucrose. The fractionated coconut oil and the aqueous solution containing dissolved HPMC and sucrose were homogenised in a high speed colloid mill, Ultra-Turrax® T25 basic (IKA Labortechnik, Germany) for 3 min at 24 000 rpm. Aqueous sucrose solutions with 50% dry powder mass, as well as aqueous mixture with 20% dry powder mass containing sucrose and HPMC at various ratios, were prepared.

The liquid o/w-emulsions and aqueous solutions of sucrose and sucrose-HPMC mixtures were spray dried in a laboratory spray dryer, Mobil Minor (Niro Atomizer, Denmark). The dryer operates co-currently, has a rotary atomizer and a flow of drying air at approximately 135 kg/h. The inlet air temperature was 120°C, the outlet air temperature was held at 75°C, and the rotation rate of the

rotary cup atomizer was 25 000 rpm. In the spray drying process, the atomizer was cooled by water.

2.3. Reconstitution

A total of 1.0 g of dry emulsion was suspended in 4.0 ml of distilled water in a 17 ml container. After 1 h of rotation at approximately 20 rpm (comfort Heto Mastermix rotator), samples were withdrawn for further characterisation.

2.4. Storage of dry emulsions, amorphous sucrose and mixtures

Dry emulsions, amorphous sucrose and amorphous sucrose-HPMC mixtures were stored at ambient temperature in desiccators over dry silica gel. Samples were taken out to be stored for 6 months at ambient temperature and 40°C in 75% relative humidity, created by a saturated NaCl solution.

2.5. Study design

A three-step study plan was set up. The first step, the reconstitution properties of the dry emulsions and the degree of crystallisation of amorphous sucrose in the dry emulsions upon ageing were investigated. The degree of crystallisation was determined by differential scanning calorimetry (DSC) analysis as the ratio between the melting enthalpy of sucrose in the dry emulsions and crystalline sucrose. The dry emulsions were heated from 25 to 225°C and the enthalpy of melting was determined as the area of the endotherm.

Subsequently, the ability of HPMC to raise $T_{\rm g}$ of amorphous sucrose-HPMC mixtures, relative to the $T_{\rm g}$ of amorphous sucrose was investigated. $T_{\rm g}$ of different sucrose-HPMC mixtures was calculated applying the Gordon-Taylor equation and measured experimentally by DSC analysis. The midpoint $T_{\rm g}$ was determined. The samples were heated from 25 to 100°C, and then kept at 100°C for 40 min to ensure dry samples (the chemical stability of sucrose was not affected). Cooled to 25°C and subsequently heated a second time to 225°C during which the midpoint $T_{\rm g}$ was determined [24,29–32]. The experiment was performed twice.

Finally the physical stability of amorphous sucrose and the dry emulsions were estimated by an enthalpy relaxation study. From the measured enthalpy recovery at $T_{\rm g}$, the mean relaxation time parameters τ and β were calculated applying the Williams-Watts equation.

By DSC analysis, the midpoint $T_{\rm g}$, the enthalpy recovery ΔH and the change in heat capacity at $T_{\rm g}$ ($\Delta C_{\rm p}$) for the samples were determined. The samples were heated from 25 to 21–22°C above $T_{\rm g}$ and then cooled to 0°C to form a sample with a standardised thermal history. The samples were then heated to a storage temperature of 20, 30 or 50°C below $T_{\rm g}$ and were held isothermally at that temperature for 1, 2, 4, 8 or 16 h. The samples were subsequently cooled and then heated through $T_{\rm g}$ to 225°C. During the final

heating scan, the pronounced endothermic relaxation peak located at the end of the glass transition region was analysed [24–25,29,31,33]. Each experiment was performed twice.

The glass transition temperature of a mixture $T_{g(mix)}$ is expressed by the Gordon-Taylor equation Eq. (1), which assumes that two components are miscible and that the free volumes of the components are additive [34]

$$T_{g(\text{mix})} = \frac{w_1 \times T_{g1} + K \times w_2 \times T_{g2}}{w_1 + K \times w_2}$$
(1)

 w_1 and w_2 are the mass fractions of each component and $T_{\rm g1}$ and $T_{\rm g2}$ are the respective glass transition temperatures. Applying free volume theory, the constant K is calculated by the Simha-Boyer rule Eq. (2) [35]

$$K \approx \frac{T_{\rm g1} \times \rho_1}{T_{\rm g2} \times \rho_2} \tag{2}$$

 ρ_1 and ρ_2 are the density of both components.

By DSC analysis an endothermic relaxation peak accompanying the glass transition is generated as a consequence of enthalpy recovery. The intensity of the peak depends on the storage conditions. The maximum enthalpy recovery ΔH_{∞} can only be determined accurately at ageing temperatures near $T_{\rm g}$ [36]. However, assuming that the heat capacity change $\Delta C_{\rm p}$ is independent of the temperature, an approximate value of ΔH_{∞} for a particular ageing temperature T can be calculated from Eq. (3)

$$\Delta H_{\infty} = (T_{\rm g} - T) \times \Delta C_{\rm p} \tag{3}$$

 $\Delta C_{\rm p}$ is the change in heat capacity at $T_{\rm g}$. From ΔH_{∞} the extent to which a material relaxes $\phi(t)$ is calculated under any given time t and temperature T conditions from Eq. (4)

$$\phi(t) = 1 - \frac{\Delta H_t}{\Delta H_{\infty}} \tag{4}$$

 $\Delta H_{\rm t}$ is the measured enthalpy recovery under those conditions and is obtained by calculating the area under the endothermic relaxation peak. The mean molecular relaxation time constant τ is calculated applying the Williams-Watts equation [37] Eq. (5)

$$\phi(t) = \exp((-t/\tau)^{\beta}) \tag{5}$$

 β is a relaxation time distribution parameter with a value between 0 and 1. A β value of unity corresponds to a single relaxation time of an exponential type.

The mean relaxation time constant τ and the relaxation time distribution parameter β are calculated from the experimental enthalpy recovery data. An iterative nonlinear regression procedure is applied to estimate the best fit to the data applying Quattro® Pro Version 7 (Corel, Canada). The initial parameters provided are $\tau=100$ and $\beta=0.5$ for all samples.

2.6. Differential scanning calorimetry (DSC)

The physical state of sucrose in the dry emulsions, amor-

phous sucrose and the sucrose-HPMC mixtures was determined by differential scanning calorimetry applying a DSC 7 equipped with an instrument controller TAC 7/PC (Perkin Elmer, Denmark) and a cooling accessory Ultra-Kryomat® Rul 80 (Lauda, Germany). A total of 10–20 mg of samples were weighed and analysed in 40 μl aluminium pans with holes and the effluent gas was dry nitrogen. The scanning rate was 20°C/min. The instrument was calibrated towards indium and zinc.

2.7. Thermogravimetric analysis (TGA)

The moisture content in the dry emulsions, amorphous sucrose and sucrose-HPMC mixtures was determined by thermogravimetric analysis (TGA) applying a TGA 7 equipped with a TAC 7/PC instrument controller (Perkin Elmer, Denmark). Samples (15.00–20.00 mg) were placed in the sample pan and the effluent gas was dry nitrogen. The scanning rate was 10°C/min in the scan range 50–200°C. The moisture content was determined as the weight loss between 50 and 120°C. The instrument was calibrated towards alumel and nickel in addition to a furnace and weight calibration.

2.8. Droplet size

The droplet size distribution of the reconstituted o/w-emulsions was determined by laser diffraction applying a Mastersizer S (Malvern Instruments Ltd., UK). A reverse Fourier optic lens with a 300 focal length was used, covering the size interval 0.05–880 µm. The refractive index of lipid was 1.4500, the absorption value was 0.1 and the refractive index of water was 1.3300. The droplet size distribution was determined on the basis of the volume distribution. The volume weighted median diameter d(v, 0.5), was applied to characterise the droplet size. The width of the droplet size distribution was expressed by the SPAN-value.

2.9. Scanning electron microscopy (SEM)

The outer macroscopic structure of the dry emulsions was examined by scanning electron microscopy applying a JMS-5200 scanning electron microscope (Jeol, Japan). Prior to microscopy, samples were coated with gold/palladium by sputtering for 300 s in a BioRad, E5200 Auto Sputter Coater. The samples were scanned at a voltage of 15 kV.

2.10. Density

The density of the dry emulsions, amorphous sucrose and HPMC was determined at ambient temperature by helium pycnometry applying an AccuPyc 1330 pycnometer (Micrometritics Ltd., UK). An equilibration rate at 0.7 kPag/min was applied and for one determination each sample was measured seven times.

Table 1
Droplet size distribution of liquid o/w-emulsions after reconstitution, the moisture content and crystallisation of amorphous sucrose in the dry emulsions^a

Ambient temperature, 75% RH					40°C, 75% RH			
Time (month)	Crystallinity (%)*	Moisture content (%)*	d(v, 0.5) (μm)	SPAN	Crystallinity (%)*	Moisture content (%)*	d(v, 0.5) (μm)	SPAN
0	_	2.0	1.71 (0.04)	1.47 (0.05)	_	2.0	1.71 (0.04)	1.47 (0.05)
1	_	1.6	1.69 (0.01)	1.43 (0.01)	_	2.0	1.67 (0.02)	1.45 (0.02)
2	_	1.5	1.64 (0.03)	1.46 (0.02)	4	2.9	1.60 (0.01)	1.47 (0.01)
3	_	1.6	1.64 (0.00)	1.53 (0.02)	13	4.0	1.63 (0.02)	1.43 (0.01)
4	_	1.8	1.68 (0.00)	1.28 (0.00)	18	5.0	1.62 (0.01)	1.43 (0.02)
5	<1	2.2	1.65 (0.02)	1.44 (0.01)	23	6.0	1.62 (0.05)	1.42 (0.06)
6	<1	2.4	1.69 (0.07)	1.51 (0.07)	25	6.0	1.67 (0.05)	1.53 (0.05)

^a The values are the mean value of two determinations and the values in brackets are the difference between the two determinations. *One determination.

3. Results

3.1. Stability of dry emulsions

Table 1 shows that during storage at 75% relative humidity, the moisture contents of the dry emulsions were increased from 2.0 to 2.4% at ambient temperature and to 6.0% at 40°C. The crystallisation of amorphous sucrose was initiated after 5 months of storage at ambient temperature and after 2 months at 40°C. At the end of the storage period the degree of crystallisation was below 1 and 25%, respectively. The reconstitution properties of the dry emulsions were not affected by the crystallisation of amorphous sucrose. The dry emulsions were able to reconstitute the original o/w-emulsion in water after 6 months of storage in 75% relative humidity at ambient temperature and 40°C. As seen in Fig. 1, the outer structure of the dry emulsions was not changed after storage in 75% relative humidity at 40°C for 6 months. The dry emulsions still consisted of well-separated spherical particles with shallow dents, seen to be deeper and more abundant in the smaller particles.

3.2. The effect of HPMC on T_g

Fig. 2 shows a plot of $T_{\rm g}$ values of sucrose-HPMC mixtures as a function of their compositions. The graph includes the values calculated by the Gordon-Taylor equation using values from Table 2. The experimental determined $T_{\rm g}$ values of the sucrose-HPMC mixtures were very close to the $T_{\rm g}$ of pure sucrose.

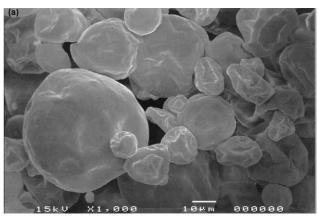
3.3. Molecular mobility below T_g

Fig. 3 shows the enthalpy recovery of amorphous sucrose and the dry emulsions as a function of ageing time at different ageing temperatures below $T_{\rm g}$. The enthalpy recovery increased with increasing ageing time and temperature in a non-linear way. The enthalpy recovery was lower for the dry emulsions even though the ageing temperatures were higher.

Fig. 4 shows the extent of relaxation of amorphous sucrose and the dry emulsions in terms of $\phi(t)$ as a function

of the ageing time at different ageing temperatures below $T_{\rm g}$. There was a significant reduction in the extent of relaxation for amorphous sucrose and the dry emulsions at temperatures 50°C below $T_{\rm g}$, and the values were almost identical. For the dry emulsions, values of °(t) were lower than for amorphous sucrose at temperatures 20 and 30°C below $T_{\rm g}$.

Table 3 shows the calculated values of the molecular relaxation time parameters τ and β based on Williams-Watts equation at different ageing temperatures. For both



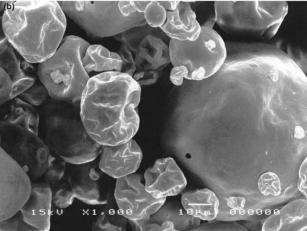


Fig. 1. Scanning electron micrograph of the dry emulsion: (a) before storage; and (b) after 6 months of storage in 75% relative humidity at 40° C. Bar = $10~\mu$ m.

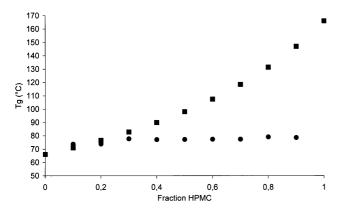


Fig. 2. $T_{\rm g}$ values of sucrose-HPMC mixtures as a function of composition: \bullet represent measured values and \blacksquare represent calculated values from the Gordon-Taylor equation.

amorphous sucrose and the dry emulsions, values of τ increased with lower ageing temperatures.

4. Discussion

Values of T_g for the various sucrose-HPMC mixtures remain equal and very close to the $T_{\rm g}$ of amorphous sucrose. T_{g} increased by 12°C relatively to amorphous sucrose with HPMC concentration up to 30% HPMC. At higher concentration deviations from the Gordon-Taylor equation were seen. This indicates a non-ideal mixing above 30% HPMC. Sucrose was shown to be miscible with HPMC during the DSC scan by a single glass transition temperature $T_{\rm g}$ and by the fact that no other thermal transitions reflecting crystallisation were observed at HPMC concentrations above 30%. Mixing HPMC with sucrose may introduce a considerable excess of free volume to the mixture due to the larger molecular size. Thus the $T_{\rm g}$ of the mixture is less elevated than calculated by Gordon-Taylor equation [29]. Crystal inhibition is obtained despite an apparent lack of antiplasticizing (i.e. T_g-elevating) effects [22], and consequently the effect of the polymer is not linked directly to $T_{\rm g}$, so other factors may be involved. For the dry emulsion containing 40% HPMC, T_g was also increased by 12°C relatively to amorphous sucrose. Dry emulsion containing 40%HPMC instead of 30% HPMC was selected due to stability

Table 2 The glass transition temperature (T_g) , the change in heat capacity (ΔC_p) at T_g and the density of the dry emulsions, amorphous sucrose and HPMC^a

Materials	T _g (°C)	$\Delta C_{\rm p} ({\rm J/g^{\circ}C})$	Density (g/cm ³)
Dry emulsions	78 ± 3.3	0.156 ± 0.002	1.20 (0.00)
Amorphous sucrose	66 ± 3.0	0.545 ± 0.002	1.53 (0.00)
HPMC	166 (0.6)	0.098 (0.000)	1.29 (0.00)

 $[^]a$ The values for $T_{\rm g}$ and $\Delta C_{\rm p}$ for the dry emulsions and amorphous sucrose are mean values of 30 determinations. The other values are the mean value of two determinations and the values in brackets are the difference between the two determinations.

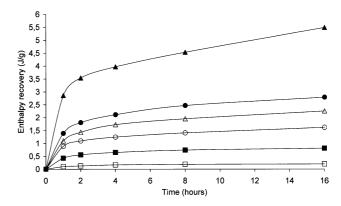


Fig. 3. Variation of enthalpy recovery (ΔH) for amorphous sucrose and the dry emulsions with time after storage at different temperatures. Ageing temperatures for amorphous sucrose: \blacksquare 17°C ($T_g - 49^{\circ}$ C), \blacksquare 37°C ($T_g - 29^{\circ}$ C) and \blacksquare 47°C ($T_g - 19^{\circ}$ C). Ageing temperatures for the dry emulsions: \square 28°C ($T_g - 50^{\circ}$ C), \square 48°C ($T_g - 30^{\circ}$ C) and \square 58°C ($T_g - 20^{\circ}$ C).

of the liquid o/w-emulsion but also to be sure that crystal inhibition of amorphous sucrose was obtained.

 β describes the distribution of molecular relaxation times. In this study values of β were different from 1, indicating that a distribution of time scales with non-exponential behaviour was required to describe each set of data, which is typically for amorphous systems.

The mean relaxation time constant τ was found to vary from a few hours near $T_{\rm g}$ to many years at temperatures 50°C below $T_{\rm g}$. The molecular mobility at 17°C for amorphous sucrose and at 28°C for the dry emulsions becomes negligible with respect to the shelf life stability. The estimated stability is in agreement with the results obtained from the conventional stability study. Below 1% amorphous sucrose in the dry emulsions was crystallised after 6 months of storage in 75% relative humidity at ambient temperature. In contrary, 25% of the amorphous sucrose was crystallised at 40°C. It is most likely that water acts as a plasticizer for sucrose. At elevated storage temperatures, the low amount of water required to reduce $T_{\rm g}$ [38] has profound implica-

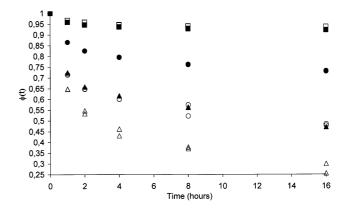


Fig. 4. The extent of relaxation for amorphous sucrose and the dry emulsions with time after storage at different temperatures. Ageing temperatures for amorphous sucrose: \blacksquare 17°C (T_g – 49°C), \bullet 37°C (T_g – 29°C) and \blacktriangle 47°C (T_g – 19°C). Ageing temperatures for the dry emulsions: \Box 28°C (T_g – 50°C), \bigcirc 48°C (T_g – 30°C) and \triangle 58°C (T_g – 20°C).

Table 3 Calculated molecular relaxation time values τ and β applying the Williams-Watts equation for amorphous sucrose and the dry emulsions at different ageing temperatures

Materials	Ageing temperatures				
		$T_{\rm g}$ – 20°C	$T_{\rm g}-30^{\circ}{\rm C}$	$T_{\rm g}$ – 50°C	
Amorphous sucrose	τ (hours)	46 0.29	1219 0.26	10^{7} 0.22	
Dry emulsions	τ (hours) β	8 0.38	51 0.27	10^{7} 0.23	

tions for accelerated stability testing of amorphous substances.

After storage for 6 months, the outer structure of the dry emulsions was not changed and the dry emulsions were able to reconstitute the original o/w-emulsion in water. The dry emulsions are physically stable with respect to the lifetime of a pharmaceutical product, when stored in dry condition and at temperatures up to 28°C.

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References

- H. Takeuchi, H. Sasaki, T. Niwa, T. Hino, Y. Kawashima, K. Uesugi, H. Ozawa, Redispersible dry emulsion system as novel oral dosage form of oily drugs: in vivo studies in beagle dogs, Chem. Pharm. Bull. 39 (1991) 3362–3364.
- [2] C.J.H. Porter, S.A. Charman, R.D. Williams, M.V. Bakalova, W.N. Charman, Evaluation of emulsifiable glasses for the oral administration of cyclosporin in beagle dogs, Int. J. Pharm. 141 (1996) 227–237.
- [3] S. Corveleyn, J.P. Remon, Bioavailability of hydrochlorothiazide: conventional versus freeze-dried tablets, Int. J. Pharm. 173 (1998) 149–155.
- [4] V.A. Richter, K. Steiger-Trippi, Untersuchungen über die zerstäubungstrocknung von emulgierten arzneizubereitungen, Pharm. Acta Helv. 36 (1961) 322–337.
- [5] M. Lladser, C. Medrano, A. Arancibia, The use of supports in the lyophilization of oil-in-water emulsions, J. Pharm. Pharmacol. 20 (1968) 450–455.
- [6] Y. Nakamoto, M. Hashida, S. Muranishi, H. Sezaki, Studies on pharmaceutical modification of anticancer agents. Enhanced delivery of bleomycin into lymph by emulsions and drying emulsions, Chem. Pharm. Bull. 23 (1975) 3125–3131.
- [7] P.E. Macheras, C.I. Reppas, Studies on drug-milk freeze-dried formulations I: bioavailability of sulfamethizole and dicumarol formulations, J. Pharm. Sci. 75 (1986) 692–696.
- [8] H. Takeuchi, H. Sasaki, T. Niwa, T. Hino, Y. Kawashima, K. Uesugi, M. Kayano, Y. Miyake, Preparation of powdered redispersible vitamin E acetate emulsion by spray-drying technique, Chem. Pharm. Bull. 39 (1991) 1528–1531.
- [9] H. Takeuchi, H. Sasaki, T. Niwa, T. Hino, Y. Kawashima, K. Uesugi,H. Ozawa, Design of redispersible dry emulsion as an advanced

- dosage form of oily drug (vitamin E nicotinate) by spray-drying technique, Drug Dev. Ind. Pharm. 18 (1992) 919–937.
- [10] H. Takeuchi, H. Sasaki, T. Niwa, T. Hino, Y. Kawashima, K. Uesugi, H. Ozawa, Improvement of photostability of ubidecarenone in the formulation of a novel powdered dosage form termed redispersible dry emulsion, Int. J. Pharm. 86 (1992) 25–33.
- [11] S.L. Myers, M.L. Shively, Preparation and characterisation of emulsifiable glasses: oil-in-water and water-in-oil-in-water emulsions, J. Colloid Interface Sci. 149 (1992) 271–278.
- [12] S.P. Vyas, C.P. Jain, A. Kaushik, V.K. Dixit, Preparation and characterisation of griseofulvin dry emulsion, Pharmazie 47 (1992) 463–464
- [13] M.L. Shively, Characterisation of oil-in-water emulsions prepared from solid-state emulsions: effect of matrix and oil phase, Pharm. Res. 10 (1993) 1153–1156.
- [14] M.L. Shively, Droplet size distribution within oil-in-water emulsions prepared from solid state dispersions, J. Colloid Interface Sci. 155 (1993) 66–69.
- [15] C. Molina, R. Cadorniga, Physical stability of lyophilized and sterilised emulsions, S.T.P. Pharma Pratiques 5 (1995) 63–72.
- [16] M.L. Shively, D.C. Thompson, Oral bioavailability of vancomycin solid-state emulsions, Int. J. Pharm. 117 (1995) 119–122.
- [17] S. Corveleyn, J.P. Remon, Formulation of a lyophilized dry emulsion tablet for the delivery of poorly soluble drugs, Int. J. Pharm. 166 (1998) 65–74.
- [18] G.P. Pedersen, P. Fäldt, B. Bergenståhl, H.G. Kristensen, Solid state characterisation of a dry emulsion: a potential drug delivery system, Int. J. Pharm. 171 (1998) 257–270.
- [19] K. Heinzelmann, K. Franke, Using freezing and drying techniques of emulsions for the microencapsulation of fish oil to improve oxidation stability, Colloids Surf. B 12 (1999) 223–229.
- [20] K.L. Christensen, G.P. Pedersen, H.G. Kristensen, Preparation of redispersible dry emulsions by spray drying, Int. J. Pharm. 212 (2001) 187–194.
- [21] K.L. Christensen, G.P. Pedersen, H.G. Kristensen, Technical optimisation of redispersible dry emulsions, Int. J. Pharm. 212 (2001) 195– 202.
- [22] S.L. Shamblin, E.Y. Huang, G. Zografi, The effects of co-lyophilized polymeric additives on the glass transition temperature and crystallisation of amorphous sucrose, J. Thermal Anal. 47 (1996) 1567– 1579.
- [23] S.L. Shamblin, L.S. Taylor, G. Zografi, Mixing behaviour of colyophilized binary systems, J. Pharm. Sci. 87 (1998) 694–701.
- [24] S.L. Shamblin, G. Zografi, Enthalpy relaxation in binary amorphous mixtures containing sucrose, Pharm. Res. 15 (1998) 1828–1834.
- [25] B.C. Hancock, S.L. Shamblin, G. Zografi, Molecular mobility of amorphous pharmaceutical solids below their glass transition temperatures, Pharm. Res. 12 (1995) 799–806.
- [26] T. Matsumoto, G. Zografi, Physical properties of solid molecular dispersions of indomethacin with poly(vinylpyrrolidone) and poly(vinylpyrrolidone-co-vinyl-acetate) in relation to indomethacin crystallization, Pharm. Res. 16 (1999) 1722–1728.
- [27] P. Tong, G. Zografi, Solid-state characteristics of amorphous sodium indomethacin relative to its free acid, Pharm. Res. 16 (1999) 1186– 1192.
- [28] G. Van den Mooter, P. Augustijns, R. Kinget, Stability prediction of amorphous benzodiazepines by calculation of the mean relaxation time constant using the Williams-Watts decay function, Eur. J. Pharm. Biopharm. 48 (1999) 43–48.
- [29] B.C. Hancock, G. Zografi, Characteristics and significance of the amorphous state in pharmaceutical systems, J. Pharm. Sci. 86 (1997) 1–12.
- [30] Y. Roos, M. Karel, Amorphous state and delayed ice formation in sucrose solutions, Int. J. Fd. Sci. Technol. 26 (1991) 553–566.
- [31] S.D. Clas, C.R. Dalton, B.C. Hancock, Differential scanning calorimetry applications in drug development, PSTT 2 (1999) 311–320.
- [32] D.Q.M. Craig, P.G. Royall, V.L. Kett, M.L. Hopton, The relevance of

- amorphous state to pharmaceutical dosage forms: glassy drugs and freeze dried systems, Int. J. Pharm. 179 (1999) 179-207.
- [33] E. Fukuoka, M. Makita, S. Yamamena, Some physicochemical properties of glassy indomethacin, Chem. Pharm. Bull. 34 (1986) 4314–4321
- [34] M. Gordon, J.S. Taylor, Ideal copolymers and the second-order transitions of synthetic rubbers. 1. Non-crystalline copolymers, J. Appl. Chem. 2 (1952) 493–500.
- [35] R. Simha, R.F. Boyer, On a general relation involving the glass temperature and coefficients of expansion of polymers, J. Chem. Phys. 37 (1962) 1003–1007.
- [36] S. Montserrat, Physical ageing studies in epoxy resins. 1. Kinetics of the enthalpy relaxation process in a fully cured epoxy resin, J. Polym. Sci. Polym. Phys. 32 (1994) 509–522.
- [37] G. Williams, D.C. Watts, Non-symmetrical dielectric relaxation behaviour arising from a simple empirical decay function, Trans. Faraday Soc. 66 (1970) 80–85.
- [38] C. Ahlneck, G. Zografi, The molecular basis of moisture effects on the physical and chemical stability of drugs in the solid state, Int. J. Pharm. 62 (1990) 87–95.